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Simultaneous determination of inorganic anions and cations by ion chromatography with indirect photometric detection

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ABSTRACT

Inorganic cations and anions were separated simultaneously on a silica-based Shim-pack WAX-1 column. The method is shown to provide efficient separation of Ca^{2+} , Mg^{2+} , Cl^- and NO_3^- . Using 1 mM phthalate-6 mM EDTA at pH 6.57 as the eluent, a mixture containing these ions was separated in less than 6 min with detection limits of 1.2-2 ng. The effects of the eluent pH, concentration and composition on the retention behaviour of the analytes were studied. Two practical applications are described.

INTRODUCTION

Ion chromatography coupled with indirect photometric detection (IPC), described by Small and Miller [1], is a simple but powerful technique for the separation and detection of many non-UV-absorbing inorganic and organic ions. A feature of this photometric approach is the use of UV-absorbing eluents that can displace the sample ions from the chromatographic column with detection in the effluent. Detection involves measuring the absorbance decrease (negative peak) as the analyte displaces a UV-active component in the mobile phase. This single-column method is at least as sensitive as the more conventional conductimetric detection method [2], and allows greater control over instrumental sensitivity. The method has been used for the separation of inorganic ions in a number of applications [3–7].

In general, anions are separated on an anionexchange resin and cations on a cation-exchange

Most of the commonly used columns in IPC are low capacity ion-exchange resins or silicabased packing exchangers. In this work, a Shimpack WAX-1 column (50×4 mm I.D.; 3 μ m)

resin. However, Fritz et al. [8] and Mou and Liu [9] reported that some transition metal ions could be separated with chloride or oxalic acid as eluent using a single anion-exchange column and a post-column derivatization detection method with 4-(2-pyridylazo)resorcinol. Matsushita [10] effected the chromatographic separation of inorganic anions and alkaline earth metal ions with EDTA as eluent, which converted metal cations into metal chelate anions using a single anionexchange column and direct UV detection in series with a conductivity detector. The use of IPC in conjunction with anion- and cation-exchange columns in series for the simultaneous determination of anions and cations but independently of each other using a single chromatograph has been described [11]. Recently, IPC for the simultaneous determination of inorganic cations and anions on a mixed bed of anion and cation exchangers was reported by Pietrzyk et al. [12].

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was selected. Surface silanol groups of the packings are chemically modified with a hydrophobic polymer to produce a neutral silica surface. A weakly basic stationary phase is chemically bonded to this silica support, producing a stable packing material for high-performance anionexchange chromatography. The ion-exchange groups introduced on to the silica surface are tertiary amino groups. However, the column conditions for the simultaneous separation of cations and anions in a single anion-exchange column and using an indirect photometric detector must be modified. This paper describes a procedure by using 1 mM phthalate-6 mM EDTA as the eluent. The effects of the eluent composition, concentration, and pH value on the retention behaviour and detection sensitivity are discussed.

EXPERIMENTAL

Instrumentation and reagents

The liquid chromatograph used consisted of a Rheodyne Model 7125 valve injector ($20-\mu l$ loop) and a Shimadzu (Kyoto, Japan) system including a solvent-delivery pump, an SPD-6AV UV–Vis spectrophotometric detector, a C-R3A Chromatopac and a Shim-pack WAX-1 column. UV spectral scans were produced with a U-3400 spectrophotometer (Hitachi).

All reagents were of analytical-reagent grade and aqueous solutions were prepared using doubly distilled water.

Chromatographic procedures

Mobile phases were prepared using analyticalreagent grade phthalic anhydride dissolved in water. The pH of each mobile phase was adjusted by dropwise addition of 1.0 M sodium hydroxide solution. All mobile phases were filtered through a 0.45- μ m filter and degassed in an ultrasonic bath prior to use. Further chromatographic conditions are given in the figure captions.

RESULTS AND DISCUSSION

The UV spectrum from 250 to 330 nm of a phthalate solution (pH 3.72) shows an absorp-

tion maximum at 280 nm with a molar absorptivity of 1279 $1/mol \cdot cm$. No other peaks are present in the spectrum. The UV spectrum from 200 to 330 nm of an EDTA solution (pH 5.56) shows no absorption. We chose 280 nm as the detection wavelength throughout this work.

The dissociation constants, pK_a , for phthalic acid (H₂P) are 2.95 and 5.41 [13]; the pK_a values for EDTA(H₄Y) are 0.9 (pK_{a_1}), 1.6 (pK_{a_2}), 2.0 (pK_{a_3}), 2.67 (pK_{a_4}), 6.16 (pK_{a_5}) and 10.26 (pK_{a_6}) [13]. It can be calculated that the solutions of phthalate and EDTA at pH 7.21 contain 98.44% P²⁻, 91.74% HY³⁻ and 8.18% H₂Y²⁻. It can be concluded from the above that P²⁻ and HY³⁻ are the predominant anions in the eluent if pH \ge 6.2. Under these conditions, the reaction between an alkaline earth metal ion M²⁺ to form a chelate anion with HY³⁻ can be written as

 $M^{2+} + HEDTA^{3-} \Longrightarrow MEDTA^{2-} + H^+$

The separation of Ca^{2+} , Mg^{2+} , Cl^{-} and NO_{3}^{-} using a solution containing 1 mM H₂P and 6 mM EDTA at pH 6.57 as the eluent is shown in Fig. 1.



Fig. 1. Separation of a mixture of cations and anions by IPC. Eluent, 1 mM H₂P-6 mM EDTA (pH 6.57); flow-rate, 0.6 ml/min; column temperature, 25°C; detection wavelength, 280 nm; absorbance units full-scale, 0.04. Peaks; $A = 0.9 \ \mu g$ Ca^{2+} ; $B = 0.5 \ \mu g Mg^{2+}$; $C = 0.8 \ \mu g Cl^{-}$; $D = 1.4 \ \mu g NO_3^{-}$.

Effect of basicity on eluent strength

In this work, the acidity of the eluent over a wide range did not affect the capacity of the strong ion-exchange column, but mainly affected the dissociation of H_2P and EDTA in the eluent. Some details are as follows.

In the anion-exchange process A^{x-} as a counter ion undergoes an ion exchange with the chelate anion and inorganic anion. The exchange process is

$$yA_{(S)}^{x-} + xB_{(M)}^{y-} \Longrightarrow yA_{(M)}^{x-} + xB_{(S)}^{y-}$$

where S and M represent the stationary and mobile phase, respectively, and A^{x-} and B^{y-} represent the counter ion and sample ion, respectively. Assuming activity coefficients to be equal to unity, we express the selectivity coefficient $K_{\rm A}^{\rm B}$ as

$$K_{A}^{B} = \frac{[A^{x^{-}}]_{(M)}^{y}[B^{y^{-}}]_{(S)}^{x}}{[A^{x^{-}}]_{(S)}^{y}[B^{y^{-}}]_{(M)}^{x}}$$
(1)

The distribution coefficient for B^{y-} is

$$D_{\rm M} = \frac{[{\rm B}^{y-}]_{({\rm S})}}{[{\rm B}^{y-}]_{({\rm M})}} = \left\{\frac{[{\rm A}^{x-}]_{({\rm S})}K_{\rm A}^{{\rm B}\,1/y}}{[{\rm A}^{x-}]_{({\rm M})}}\right\}^{y/x}$$
(2)

On the other hand, an ion-exchange equilibrium between OH^- and A^{x-} also exists:

$$\mathbf{A}_{(S)}^{x-} + \mathbf{x}\mathbf{OH}_{(M)}^{-} \rightleftharpoons \mathbf{A}_{(M)}^{x-} + \mathbf{x}\mathbf{OH}_{(S)}^{-}$$

Similarly, the selectivity coefficient K_A^{OH} is

$$K_{\rm A}^{\rm OH} = \frac{[{\rm A}^{x^{-}}]_{({\rm M})}[{\rm OH}^{-}]_{({\rm S})}^{x}}{[{\rm A}^{x^{-}}]_{({\rm S})}[{\rm OH}^{-}]_{({\rm M})}^{x}}$$
(3)

By combination of eqns. 2 and 3:

$$D_{\rm M} = \frac{[{\rm B}^{y-}]_{(\rm S)}}{[{\rm B}^{y-}]_{(\rm M)}} = \frac{K_{\rm A}^{\rm B\,1/x}[{\rm OH}^{-}]_{(\rm S)}^{y}}{K_{\rm A}^{\rm OH\,y/x}[{\rm OH}^{-}]_{(\rm M)}^{y}}$$
(4)

From eqn. 4, we concluded that the higher the basicity of the eluent, the weaker is the retention of the sample ions. This is consistent with the experimental results shown in Fig. 2.

In Fig. 2, the peaks for the first three anions are completely overlapped at pH 5.30, the peaks for Mg^{2+} and Cl^- are completely overlapped in the pH range 5.30–5.94 and the peaks for Ca^{2+} and Cl^- are completely overlapped below pH



Fig. 2. Retention time versus the pH of the eluent. Eluent, 1 mM H,P-6 mM EDTA. Other conditions as in Fig. 1.

5.30. However, NO_3^- was well resolved, but the retention time for NO_3^- was extremely long at pH <5.30. It was noticeable that the retention time for Mg^{2^+} increased with increasing basicity of the eluent at pH <5.30. We presumed that MgY^{2^-} was dissociated below pH 5.3. According to the conditional equilibrium constants [14], 4.1 for CaY²⁻ and 2.1 for MgY²⁻ at pH 5.0, the stability of CaY²⁻ is 100 times higher than that of MgY²⁻, and the chelate MgY²⁻ cannot exist stably in this pH range.

The effects of pH on the retention behaviour of Cl⁻ and NO₃⁻ were expected to be dependent on the acid dissociation constants of H₂P and EDTA. However, the effects of pH on the retention behaviour of Ca²⁺ and Mg²⁺ were expected to be dependent on both the conditional stability constants of metal EDTA chelates and the acid dissociation constants of H₂P and EDTA.

Effect of basicity of eluent on detection sensitivity

Hydroxyl ions in a weakly acidic eluent can act as counter ions when the eluent pH is much greater than its pK_a value. Both A^{x-} and OH^- , which are retained by the anion exchanger, can be displaced by the sample ions. This makes the sensitivity of detection lower than the molar absorptivity of phthalate. Hence a pH of the eluent generally maintained in the acid range will produce a powerful separation and a more sensi-



Fig. 3. Plot of sample peak area versus eluent pH. Conditions as in Fig. 1.

tive detection. The plot in Fig. 3 verified the point that the detection sensitivity decreases with increasing hydroxyl ion concentration using a weakly acidic mobile phase with $pH > pK_a$ with the IPC method described in this paper.

The result indicates that the ideal eluent pH for complete and optimum separation of the four analytes should be controlled in the range 6.2–7.0; this would eliminate the deleterious effect on detection caused by hydroxyl ions and could avoid damage to the chromatographic column.

Effect of eluent concentration on signal-to-noise ratio

The peak area or peak height for analytes depends on the concentration of the IPC eluent (Fig. 4). Low eluent concentrations are recommended for achieving low background absorb-



Fig. 4. Plot of sample peak area *versus* eluent concentration. Conditions as in Fig. 1.

ance and good detection limits [1]. This was also predicted by Yeung [15] using the equation

$$C_{\rm lim} = C_{\rm m} / (D_{\rm R} T_{\rm R}) \tag{5}$$

where C_{\lim} is the concentration limit of detection, $C_{\rm m}$ is the concentration of the eluent, $D_{\rm R}$ is the dynamic reserve (ratio of background signal to background noise) and $T_{\rm R}$ is the transfer ratio, which is expressed as the number of eluent molecules displaced by one analyte molecule. Eqn. 5 shows a direct proportionality between C_{lim} and C_{m} . These trends are typical and consistent with all indirect detection techniques, because the relative change in the baseline absorbance is more pronounced at low than at high background absorbance levels. A low eluent concentration provides high sensitivity and powerful separation compared with more concentrated eluents. However, it is important to note that a too dilute eluent will result in overlong run times and a decrease in sensitivity due to band spreading.

Effect of eluent composition on retention behaviour

The elution mechanism of IPC using a mixed eluent as in this work is complicated. A series of mixed eluents containing various concentrations of H₂P and EDTA at pH 7.4 were studied, and also two single eluents, H₂P (1 mM, pH 6.95) and EDTA (6 mM, pH 7.02). Fig. 5 illustrates the chromatogram of Ca^{2+} , Mg^{2+} , Cl^{-} and NO_{3}^{-} using EDTA as eluent with detection at 230 nm. All the analytes were well resolved, but baseline noise cannot be deleted. In addition, no peaks appeared when the wavelength was set at 280 nm using the same eluent, because EDTA and the EDTA chelate do not absorb at this wavelength. Fig. 6 shows a chromatogram of the samples obtained using H₂P as eluent with detection at 280 nm. Only two negative peaks for Cl⁻ and NO_3^- were observed, and peaks for Ca^{2+} and Mg^{2+} were not found because Ca^{2+} and Mg^{2+} cations could not be retained by the anion-exchange column. The experiment was carried out by injecting MY^{2-} into the chromatographic column and two negative peaks were obtained.

From the above experiments, two theoretical



Fig. 5. Separation of a mixture of cations and anions. Eluent, 6 mM EDTA (pH 7.02); detection wavelength, 230 nm.

models were presumed. The separation of Ca^{2+} and Mg^{2+} was probably based on two factors: the competitive formation balance between metal cations and EDTA and the anion-exchange balance between EDTA anion chelate



Fig. 6. Separation of chloride and nitrate. Eluent, 1 mM H_2P (pH 6.95); detection wavelength, 280 nm.

and counter ions. When HY^{3-} and P^{2-} displaced MY^{2-} from the column into the eluent, the H_2P concentration was changed, which resulted in a negative absorption at 280 nm, corresponding to an indirection detection mode. The mechanism can be explained as follows:

$$3MY_{(M)}^{2^{-}} + 2HY_{(S)}^{3^{-}} \Longrightarrow 3MY_{(S)}^{2^{-}} + 2HY_{(M)}^{3^{-}}$$
 (6)

$$MY_{(M)}^{2^{-}} + P_{(S)}^{2^{-}} \Longrightarrow MY_{(S)}^{2^{-}} + P_{(M)}^{2^{-}}$$
 (7)

By combining eqns. 6 and 7 we obtain

$$4MY_{(M)}^{2^{-}} + 2HY_{(S)}^{3^{-}} + P_{(S)}^{2^{-}}$$

$$\implies 4MH_{(S)}^{2^{-}} + 2HY_{(M)}^{3^{-}} + P_{(M)}^{2^{-}}$$
(8)

The selectivity coefficient $K_{Y,P}^{MY}$ is

$$K_{Y,P}^{MY} = \frac{[MY^{2^{-}}]_{(S)}^{4}[HY^{3^{-}}]_{(M)}^{2}[P^{2^{-}}]_{(M)}}{[MY^{2^{-}}]_{(M)}^{4}[HY^{3^{-}}]_{(S)}^{2}[P^{2^{-}}]_{(S)}}$$
(9)

The molar fraction of MY^{2-} in solution is

$$\Phi = \frac{[MY^{2^{-}}]}{[M^{2^{+}}] + [MY^{2^{-}}]}$$
(10)

The distribution coefficient for M^{2+} can be deduced by combining eqns. 9 and 10:

$$D_{\rm M} = \frac{[{\rm M}{\rm Y}^{2^-}]_{(\rm S)}}{[{\rm M}^{2^+}]_{(\rm M)}}$$
$$= K_{\rm Y,P}^{\rm MY\,1/4} \frac{\Phi}{1-\Phi} [{\rm H}{\rm Y}^{3^-}]_{(\rm S)}^{1/2} [{\rm P}^{2^-}]_{(\rm S)}^{1/4}$$
$$\times ([{\rm H}{\rm Y}^{3^-}]_{(\rm M)}^2 [{\rm P}^{2^-}]_{(\rm M)})^{-1/4}$$
(11)

The logarithmic form of eqn. 11 is

$$\log D_{\rm M} = \frac{1}{4} \log K_{\rm Y,P}^{\rm MY} + \log \left(\frac{\Phi}{1-\Phi}\right) + \frac{1}{2} \log \left[\rm{HY}^{3-}\right]_{(S)} + \frac{1}{4} \log \left[\rm{P}^{2-}\right]_{(S)} - \frac{1}{4} \log \left(\left[\rm{HY}^{3-}\right]_{(M)}^{2} \left[\rm{P}^{2-}\right]_{(M)}\right)$$
(12)

where $K_{Y,P}^{MY}$, $[HY^{2^-}]_{(S)}$ and $[P^{2^-}]_{(S)}$ are constants. However, Φ can also be a constant if the concentration of EDTA in each of the eluents is much higher than that of metal cations. In this event, a straight line should be obtained on

plotting log K' vs. $\log\{[HY^{3-}]^2_{(M)}[P^{2-}]_{(M)}\}$, and the theoretical slope value should be -0.25.

Experiments showed that the correlation coefficients for Ca^{2+} and Mg^{2+} are -0.9994 and -0.9999, respectively, and the actual slope values for both are exactly the correct value, -0.25. Apparently the actual counter ions in the eluent for Ca^{2+} and Mg^{2+} are HY^{3-} and P^{2-} , which are compatible with the hypotheses described above.

The separation model for monovalent anions can be described as follows. It was assumed that Cl^- and NO_3^- were displaced from the column by the counter ions HY^{3-} and P^{2-} , which also resulted in a negative absorption at 280 nm, corresponding to an indirect detection mode:

$$HY_{(s)}^{3-} + 3L_{(M)}^{-} \Longrightarrow HY_{(M)}^{3-} + 3L_{(s)}^{-}$$
(13)

$$P_{(S)}^{2-} + 2L_{(M)}^{-} \Longrightarrow P_{(M)}^{2-} + 2L_{(S)}^{-}$$
(14)

By combining eqns. 13 and 14, we obtain

$$HY_{(S)}^{3-} + P_{(S)}^{2-} + 5L_{(M)}^{-}$$

$$\implies HY_{(M)}^{3-} + P_{(M)}^{2-} + 5L_{(S)}^{-}$$
(15)

The logarithmic form of $D_{\rm M}$ for L^- can be deduced similarly:

$$\log D_{\rm M} = \frac{1}{5} \log K_{\rm Y,P}^{\rm L} + \frac{1}{5} \log [{\rm P}^{2^-}]_{\rm (S)} + \frac{1}{5} \log [{\rm H}{\rm Y}^{3^-}]_{\rm (S)} - \frac{1}{5} \log ([{\rm H}{\rm Y}^{3^-}]_{\rm (M)} [{\rm P}^{2^-}]_{\rm (M)})$$
(16)

where $K_{Y,P}^{L}$, $[P^{2^{-}}]_{(S)}$ and $[HY^{3^{-}}]_{(S)}$ are constants. A linear relationship should exist between log D_{M} and log $\{[HY^{3^{-}}]_{(M)}[P^{2^{-}}]_{(M)}\}$, and the theoretical slope value should be -0.20.

Experiments indicated that the correlation coefficients for Cl⁻ and NO₃⁻ are -0.9952 and -0.9963, respectively, and the actual slope values are -0.22 and -0.20, respectively.

Six monovalent anions were separated under the same conditions and their slope values and correlation coefficients are given in Table I.

The binary eluent used is effective in separating some other common cations and anions, and the absolute retention times for these ions are given in Table II.

TABLE I

SLOPE VALUES AND CORRELATION COEFFICIENTS FOR ANIONS

Eluent, 1 mM H₂P-6 mM EDTA (pH 7.03); injection volume, 10 μ l (1 mM). Other conditions as in Fig. 1.

Anion	Slope	Correlation coefficient	
BrO ₃	-0.20	-0.9960	
Cl⁻́	-0.20	-0.9968	
NO,	-0.19	-0.9961	
CIO	-0.19	-0.9951	
Br ⁻	-0.19	-0.9937	
NO_3^-	-0.24	-0.9793	

Applications

The correlation coefficients of the calibration graphs for Ca^{2+} , Mg^{2+} , and Cl^- are excellent, varying from 0.9992 to 0.9995. The detection limits for Ca^{2+} , Mg^{2+} and Cl^- are 1.2, 1.4, and 2 ppm, respectively (signal-to-noise ratio = 2) when using the optimized mobile phase.

The IPC method described was used to determine alkaline earth metal ions and chloride in tap water and Yellow River water. The concentrations of these three ions determined by this technique are given in Table III.

TABLE II

ABSOLUTE RETENTION TIMES FOR CATIONS AND ANIONS

Ion	Absolute retention time (min)	Ion	Absolute retention time (min)
Ba ²⁺	3.08°	BrO,	2.37 ^b
Pb ²⁺	3.56°	Cl	3.02
Ni ²⁺	7.05 ^a	NO ₂	3.67 ^b
Mn ²⁺	4.81 [°]		4.00
Co ²⁺	6.48 ^a	Br ⁻	5.12 ^b
Zn^{2+}	6.65°	NO ₁	5.65 ^b
Cu ²⁺	14.86 ^a	- 3	
Bi ³⁺	3.33"		
Rh ³⁺	3.34 ^a		

^a Eluent, 1.2 mM H₂P-1.2 mM EDTA (pH 5.54); injection volume, 10 µl (1 mM). Other conditions as in Fig. 1.

^b Eluent, 1.2 mM H₂P-1.2 mM EDTA (pH 7.02); injection volume, 10 μl (1 mM). Other conditions as in Fig. 1.

TABLE III

ANALYTICAL RESULTS FOR TWO WATER SAMPLES

Eluent, 1 mM H_2P-6 mM EDTA (pH 7.25); injection volume, 20 μ l.

Sample	Ca ²⁺ (ppm) ^a	Mg ²⁺ (ppm) ^a	Cl⁻ (ppm)"
Tap water	72.0	22.6	43.2
Yellow River water	50.5	16.7	35.0

" Values are (w/w).

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